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Mini Review

Targeting adipocyte apoptosis: A novel strategy for obesity therapy

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ABSTRACT

Obesity is an increasing world problem that may cause several metabolic complications including insulin resistance, hyperlipidemia, hypertension, and atherosclerosis. Development of therapeutic drugs for obesity has been proven difficult. Current strategies for weight reduction are inhibition of food intake through the central nervous system or blocking the absorption of lipids in the gut. These therapies have many side effects, so new treatments are urgently needed. Fat loss could also be achieved through a decrease in the size and number of adipocytes through apoptosis. Apoptosis is a normal phenomenon of cell death for the purpose of maintaining homeostasis. Induction of apoptosis is a reasonable way to remove adipocytes in obese patients. It is reported that several adipokines and natural products play roles in induction of adipocyte apoptosis. Here we review the recent progress of the roles and mechanisms of adipocyte apoptosis induced by leptin, tumor necrosis factor- α (TNF- α), and natural compounds.

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1. Introduction

Obesity is a worldwide problem with rapidly increasing prevalence [1], attracting public health attention especially in Western countries [2]. Obesity is as a result of the excess calories being stored as triglyceride in adipose tissue and ectopically in other tissues [3–5], and is associated with insulin resistance, type 2 diabetes, cardiovascular disease, hypertension, hyperlipidemia, non-alcoholic steatohepatitis, stroke, and certain cancers [4,6,7]. Prevention and treatment of obesity will prevent or greatly benefit the treatment of these related diseases. However, several therapeutic drugs to treat obesity have been withdrawn from the market in the last decade [8]. Currently there is only one approved drug, orlistat, for long-term use in the treatment of obesity and new therapeutic approaches are urgently required for treatment of obesity [6].

Obesity is characterized by the increase in adipocyte size and number. Therefore, targeting adipose tissue has become a new strategy for obesity treatment, and includes suppression of adipogenesis or adipose tissue mass, obstruction of lipid accumulation in adipocytes, and adipocyte deletion by apoptosis [9–12].

Apoptosis is characterized by plasma membrane blebbing, cytoplasm condensation, DNA fragmentation, and phagocytosis of the apoptotic body by macrophages [13], all regulated by the antiapoptotic and pro-apoptotic proteins. Apoptosis is necessary for maintaining homeostasis by removing dangerous and unnecessary

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cells [14–16]. For example, anti-cancer drugs inhibit the growth of carcinoma cells and other overgrown cells through the activation of apoptosis. However, adipocytes are resistant to apoptosis because of high levels of Akt/protein kinase B and the anti-apoptotic factor Bcl-2. Adipocytes could be removed through apoptotic mechanisms in some pathological conditions, such as in patients with tumor cachexia and HIV-infected patients receiving antiretroviral therapy. Therefore, the induction of apoptosis in adipocytes could be an attractive method to reduce the adipocyte number.

2. Adipokines

As an endocrine organ, adipose tissue secretes abundant adipokines that affect the regulation of fat weight and homeostasis. These proteins build a multitudinous array of adipose targets for drugs to treat and prevent obesity [17]. Since the discovery of leptin in 1994, many attempts have been made to use it in anti-obesity therapy [18,19]. There is evidence that administration of leptin can induce adipocyte apoptosis [20].

2.1. Leptin

Leptin, a product of the obese(ob) gene, includes 167 amino acid peptides [21], is produced and secreted by adipocytes, and mediates adipose tissue mass by elevating thermogenesis and restricting food intake through both central and peripheral mechanisms [22,23]. Recent study has suggested that after intra-cerebroventricular (ICV) administration of leptin in rats, adipose tissue showed a rapid decrease and features of apoptosis. Interestingly, the apoptosis was not observed in control and pair-fed rats and in other tissues

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of leptin-treated rats [24]. Peripheral infusion of leptin is efficacious in inducing adipocyte apoptosis in mice [25]. High-fat feeding has a gender-specific role with only males, on reduced responsiveness to leptin-induced adipocyte apoptosis. Leptin does not act directly on adipocytes to induce apoptosis. The mechanism of this process is still uncertain.

Many efforts have been made to discover the mechanisms of leptin involvement in adipocyte apoptosis. It has been reported that leptin induces the expression of angiopoietin-2 in adipose tissue without an accompanying elevation in vascular endothelial growth factor (VEGF), and leads to apoptosis in adipose endothelial cells [26]. Qian and colleagues have found that PPAR- γ mRNA levels showed a 70–80% increase after ICV administration of leptin, suggesting that PPAR- γ may be involved in leptin-induced adipocyte apoptosis [27]. This result was supported by the finding that thiazolidinedione (TZD), an agonist of PPAR- γ , can evoke adipocyte differentiation and apoptosis in white adipose tissue from rodents, contributing to prevention of adipocyte hypertrophy [28]. Loss of adipose tissue through leptin-induced apoptosis may present an effective mean of prevention and treatment of obesity.

2.2. TNF-α

Patients with cancer, AIDS, and chronic obstructive lung disease may have cachexia, characterized by loss of body mass and adipose tissues. In these patients, the inflammatory cytokine TNF- α , also called *cachexin*, was significantly increased. There is evidence that TNF- α may play a role in the loss of adipocytes.

TNF- α is synthesized and produced by adipocytes and plays a crucial role in regulation and control of adipose mass and adipocyte number via apoptosis mechanisms [29,30]. A study on brown adipocytes suggests that TNF-α induces the apoptosis of differentiated brown fat cells, and this effect is possibly adjusted mainly by the P55 TNF- α receptor subtype [31]. A further study has shown that brown adipocyte apoptosis in obese mice is abated with the absence of TNF- α P55 receptor [32]. During TNF- α -induced antiadipogenesis experiment, TNF-α-induced DNA fragmentation was significantly augmented in the β-catenin knockdown preadipocytes, suggesting that a β-catenin signaling pathway may be involved in TNF-α regulation of 3T3-L1 preadipocyte apoptosis [33]. TNF- α also induces and enhances apoptosis in human preadipocytes and adipocytes [34]. A recent study shows that TNF- α reduces the number of mature adipocytes, but not preadipocytes, through the induction of apoptosis resulting in C/EBP and PPAR- γ -mediated suppression of NF-κB [35].

3. Natural compounds

Several compounds purified from plants might be effective methods for inducing adipocyte apoptosis.

3.1. Phenols

Phenols, which are commonly found in plants, have pharmacological and biochemical effects on anti-inflammatory and anti-cancer therapy [36–38]. Recent studies suggest that phenols also have anti-obesity effects. Curcumin, the major polyphenol in turmeric spice, has been shown to cause 3T3-L1 adipocyte apoptosis [39]. Green tea (GT) and GT components, particularly catechins, could induce preadipocyte differentiation and adipocyte apoptosis [40,41]. Green tea treatment also showed a high apoptotic rate in visceral adipose tissue in rats that was closely related to augmented aromatase expression, and circulating concentration of 17 β -estradiol, and plasma concentration of testosterone [42]. Resveratrol, a naturally occurring phytoalexin derived from red wines

and grape juice, induced apoptosis in 3T3-L1 adipocytes [43]. Resveratrol also induced the apoptosis of rat primary adipocytes by increasing the expression of Sirt1, Cytochrome *c*, cleaved Caspase 9, and cleaved Caspase 3 that are related to cell apoptosis [44]. Combined treatment of resveratrol, genistein, and quercetin enhanced apoptosis in pre- and lipid-filled mature murine adipocytes and in early- and mid-phase maturing and lipid-filled mature human adipocytes more than the responses to genistein, quercetin, and resveratrol when used separately [45]. Further, combination of resveratrol and genistein has a stronger effect on induction of adipocyte apoptosis [46]. Therefore, the enhanced effects due to combination of two or more natural compounds may have more potential than the individual compounds in preventing obesity.

3.2. Flavonoids

Flavonoids have been shown to have a broad spectrum of biological effects, including anti-inflammatory, anti-cancer, antioxidation, and cardiovascular maintenance effects [47]. In recent years, flavonoids have been shown to play a role in anti-obesity activity via the induction of apoptosis. Xanthohumol and isoxanthohumol could induce apoptosis in 3T3-L1 adipocytes through the activation of mitochondrial pathway and caspase-3/7 [48]. Recent study has shown that xanthohumol enhanced apoptosis in both preadipocytes and mature adipocytes. In addition, xanthohumol-treated preadipocytes show the increase of NF-κB expression that is associated with regulation of numerous cells apoptosis, differentiation, and proliferation [49,50]. Furthermore, xanthohumol plus honokiol reinforced induction of apoptosis in 3T3-L1 mature adipocytes via the increase of cleaved PARP and Cytochrome c release, and inhibition of Bcl-2 protein levels. In addition, they could activate PTEN and inhibit AKT signaling [51]. Polymethoxy flavones (PMFs), which exist exclusively in Citrus genus, have a fascinating and broad spectrum of bioactivity consisting of inducing apoptosis activity in various cell types [52–54]. In mature 3T3-L1 adipocytes, PMFs could induce apoptosis via activation of Ca²⁺-dependent calpain and Ca²⁺/calpain-dependent caspase-12. And apoptosisinducing activity of hydroxylated-PMFs is significantly higher, which may be attributed to their stimulating effect on the production of reactive oxygen species (ROS) [55]. Recent study has shown that (2S,3S)-aromadendrin-6-C-β-D-glucopyranoside (AG), a novel flavonol isolated from *Ulmus wallichiana*, also evokes apoptosis of differentiated 3T3-L1 adipocytes [56].

Isoflavones from soybean products have several biological effects, including estrogenic and hypolipidemic activities. For example, genistein has been shown to reduce body weight via induction of apoptosis of adipose tissues in ovariectomized mice [10]. Administration of genistein and daidzein has also shown increased apoptosis through the alteration of the apoptosis-related proteins including augmentation of Bad and reduction of pAkt protein [57]. The interaction between genistein and 1,25-dihydroxyvitamin D3 (1,25(OH)2D3) (vitamin D) evoked apoptosis in mature 3T3-L1 adipocytes via the increase of VDR (vitamin D receptor) [58]. These data suggest the potential of flavonoids as a therapy for obesity.

4. Other natural compounds

In addition to phenols and flavonoids, several other natural compounds have been proven to have anti-obesity activity. Esculetin (6,7-dihydroxy-2*H*-1-benzopyran-2-one) a coumarin derivative found in various plants, has been shown to enhance adipocyte apoptosis and induce apoptosis in the late stage of differentiation [59]. Ajoene, from garlic, induces apoptosis of 3T3-L1 adipocytes through generation of hydrogen peroxide, activation of mitogen-activate protein kinases, cleavage of DNA, translocation of AIF (apoptosis-

inducing factor), and reduction of PARP-1 [60]. Guggulsterone (GS), the active substance in guggulipid, may exert anti-obesity effects by evoking apoptosis in mature adipocytes through the increase of caspase-3 activity and release of Cytochrome c from mitochondria. And cis-GS is more potent than trans-GS in the induction of adipocyte apoptosis [61]. Several anti-obesity components purified from t Rubia t cordifolia t. have been reported, including mollugin and t carbomethoxy-2,3-epoxy-3-prenyl-1,4-naphthoquinone (CMEP-NQ), which belong to anthraquinones and naphthoquinones. In t T3-L1 preadipocytes treated by mollugin and CMEP-NQ have shown the appearance of apoptosis, including mitochondrial membrane potential loss, and subsequent activation of caspases cascade including caspase-9, -3, and -7, resulting in PARP cleavage [12,62,63].

From the data above, the natural compounds show the ability to evoke apoptosis in adipocytes, which may have a major role in anti-obesity therapy.

5. Biochemical components

In addition to natural compounds, there are biochemical components for preventing obesity by inducing apoptosis. For example, preadipocyte differentiation may be augmented by fatty acids in the presence of adequate hormone cocktails. Palmitate, a longchain saturated fatty acid, can provoke apoptosis in mouse 3T3-L1 and rat primary preadipocytes in the absence of adipogenic stimuli, which due to create multiple cellular stresses [64]. Docosahexaenoic acid (DHA, C22:6), a (n-3) polyunsaturated fatty acid in fish oil [65], and 1,25-dihydroxyvitamin D₃, the metabolic product of vitamin D, have been reported to have anti-obesity activity due to their roles in stimulating adipocyte apoptosis [66,67]. Clenbuterol, a selective β2-AR agonist, has been proven to induce white adipose tissue apoptosis in mice [68]. It has also been demonstrated that activation of \(\beta 2\)-adrenergic receptor by clenbuterol evokes apoptosis in adipocytes though lipolysis stimulation [69]. Therefore, biochemical compounds may present an additional option for body fat loss through apoptosis.

Development of anti-obesity drugs remains difficult, but targeting adipocyte apoptosis may provide an opportunity for the treatment and prevention of obesity. Both natural products and biological products have apoptosis-inducing activities. Adipokines such as leptin and TNF- α are important anti-obesity drug candidates. Some natural compounds also induce apoptosis and may play a major therapeutic role in weight-reduction treatment. Before pharmaceutical interventions can be marketed, there are problems to overcome. First, provoking apoptosis in adipose tissue physiologically is likely to be harmful to other tissues, just as chemotherapy drugs induce apoptosis in cancer cells but also cause off-target effects on other organs and tissues. Developing a drug that specifically targets adipose tissue is a challenge. Secondly, targeting adipocytes may cause the cells to release lipids into blood or induce the storage of lipids in other tissues such as liver and muscle. An ideal apoptosis-inducing anti-obesity drug should avoid such side effects. Finally, the apoptosis-inducing agents may cause DNA damage and increase carcinogenesis. The anti-obesity drugs should be safe for long term use.

References

- [1] M.M. Finucane, G.A. Stevens, M.J. Cowan, G. Danaei, J.K. Lin, C.J. Paciorek, G.M. Singh, H.R. Gutierrez, Y. Lu, A.N. Bahalim, F. Farzadfar, L.M. Riley, M. Ezzati, National, regional, and global trends in body-mass index since 1980: systematic analysis of health examination surveys and epidemiological studies with 960 country-years and 9.1 million participants, Lancet 377 (2011) 557–567.
- [2] Obesity: preventing and managing the global epidemic. Report of a WHO consultation, WHO Consultation. Technical Report Series 894 (2000) i–xii, 1– 253

- [3] N.G. Popovich, O.B. Wood, Drug therapy for obesity: an update, J. Am. Pharm. Assoc. (Wash.) NS37 (1997) 31–39. 56.
- [4] G. Reaven, F. Abbasi, T. McLaughlin, Obesity, insulin resistance, and cardiovascular disease, Recent Prog. Horm. Res. 59 (2004) 207–223.
- [5] M. Lafontan, Fat cells: afferent and efferent messages define new approaches to treat obesity, Annu. Rev. Pharmacol. Toxicol. 45 (2005) 119–146.
- to treat obesity, Annu. Rev. Pharmacol. Toxicol. 45 (2005) 119–146.
 [6] T.L. Visscher, J.C. Seidell, The public health impact of obesity, Annu. Rev. Public Health 22 (2001) 355–375.
- [7] A.R. Saltiel, C.R. Kahn, Insulin signalling and the regulation of glucose and lipid metabolism, Nature 414 (2001) 799–806.
- [8] J. Kolanowski, A risk-benefit assessment of anti-obesity drugs, Drug Saf. 20 (1999) 119–131.
- [9] A. Sorisky, R. Magun, A.M. Gagnon, Adipose cell apoptosis: death in the energy depot, Int. J. Obes. Relat. Metab. Disord. 24 (Suppl. 4) (2000) S3–S7.
- [10] H.K. Kim, C. Nelson-Dooley, M.A. Della-Fera, J.Y. Yang, W. Zhang, J. Duan, D.L. Hartzell, M.W. Hamrick, C.A. Baile, Genistein decreases food intake, body weight, and fat pad weight and causes adipose tissue apoptosis in ovariectomized female mice, J. Nutr. 136 (2006) 409–414.
- [11] C. Couillard, P. Mauriege, P. Imbeault, D. Prud'homme, A. Nadeau, A. Tremblay, C. Bouchard, J.P. Despres, Hyperleptinemia is more closely associated with adipose cell hypertrophy than with adipose tissue hyperplasia, Int. J. Obes. Relat. Metab. Disord. 24 (2000) 782–788.
- [12] Y. Jun do, C.R. Han, M.S. Choi, M.A. Bae, M.H. Woo, Y.H. Kim, Effect of mollugin on apoptosis and adipogenesis of 3T3-L1 preadipocytes, Phytother. Res. 25 (2011) 724-731.
- [13] I. Budihardjo, H. Oliver, M. Lutter, X. Luo, X. Wang, Biochemical pathways of caspase activation during apoptosis, Annu. Rev. Cell Dev. Biol. 15 (1999) 269-
- [14] S.H. Kaufmann, M.O. Hengartner, Programmed cell death: alive and well in the new millennium, Trends Cell Biol. 11 (2001) 526–534.
- [15] H. Everett, G. McFadden, Apoptosis: an innate immune response to virus infection, Trends Microbiol. 7 (1999) 160–165.
- [16] D.R. Green, Apoptotic pathways: paper wraps stone blunts scissors, Cell 102 (2000) 1–4.
- [17] O. Boss, N. Bergenhem, Adipose targets for obesity drug development, Expert Opin. Ther. Targets 10 (2006) 119–134.
- [18] C. Bjorbaek, B.B. Kahn, Leptin signaling in the central nervous system and the periphery, Recent Prog. Horm. Res. 59 (2004) 305–331.
- [19] H.E. Bays, Current and investigational antiobesity agents and obesity therapeutic treatment targets, Obes. Res. 12 (2004) 1197–1211.
- [20] P.S. Gullicksen, M.A. Della-Fera, C.A. Baile, Leptin-induced adipose apoptosis: implications for body weight regulation, Apoptosis 8 (2003) 327–335.
- [21] P. Prolo, M.L. Wong, J. Licinio, Leptin, Int. J. Biochem. Cell Biol. 30 (1998) 1285– 1290
- [22] Y. Zhang, R. Proenca, M. Maffei, M. Barone, L. Leopold, J.M. Friedman, Positional cloning of the mouse obese gene and its human homologue, Nature 372 (1994) 425–432.
- [23] R.V. Considine, J.F. Caro, Leptin and the regulation of body weight, Int. J. Biochem. Cell Biol. 29 (1997) 1255–1272.
- [24] H. Qian, M.J. Azain, M.M. Compton, D.L. Hartzell, G.J. Hausman, C.A. Baile, Brain administration of leptin causes deletion of adipocytes by apoptosis, Endocrinology 139 (1998) 791–794.
- [25] M.A. Della-Fera, C. Li, C.A. Baile, Resistance to IP leptin-induced adipose apoptosis caused by high-fat diet in mice, Biochem. Biophys. Res. Commun. 303 (2003) 1053–1057.
- [26] B. Cohen, D. Barkan, Y. Levy, I. Goldberg, E. Fridman, J. Kopolovic, M. Rubinstein, Leptin induces angiopoietin-2 expression in adipose tissues, J. Biol. Chem. 276 (2001) 7697–7700.
- [27] H. Qian, G.J. Hausman, M.M. Compton, M.J. Azain, D.L. Hartzell, C.A. Baile, Leptin regulation of peroxisome proliferator-activated receptor-gamma, tumor necrosis factor, and uncoupling protein-2 expression in adipose tissues. Biochem. Biophys. Res. Commun. 246 (1998) 660-667.
- [28] T. Yamauchi, J. Kamon, H. Waki, K. Murakami, K. Motojima, K. Komeda, T. Ide, N. Kubota, Y. Terauchi, K. Tobe, H. Miki, A. Tsuchida, Y. Akanuma, R. Nagai, S. Kimura, T. Kadowaki, The mechanisms by which both heterozygous peroxisome proliferator-activated receptor gamma (PPARgamma) deficiency and PPARgamma agonist improve insulin resistance, J. Biol. Chem. 276 (2001) 41245–41254.
- [29] P.A. Kern, M. Saghizadeh, J.M. Ong, R.J. Bosch, R. Deem, R.B. Simsolo, The expression of tumor necrosis factor in human adipose tissue. Regulation by obesity, weight loss, and relationship to lipoprotein lipase, J. Clin. Invest. 95 (1995) 2111–2119.
- [30] J.K. Sethi, G.S. Hotamisligil, The role of TNF alpha in adipocyte metabolism, Semin. Cell Dev. Biol. 10 (1999) 19–29.
- [31] E. Nisoli, L. Briscini, C. Tonello, C. De Giuli-Morghen, M.O. Carruba, Tumor necrosis factor-alpha induces apoptosis in rat brown adipocytes, Cell Death Differ. 4 (1997) 771–778.
- [32] E. Nisoli, L. Briscini, A. Giordano, C. Tonello, S.M. Wiesbrock, K.T. Uysal, S. Cinti, M.O. Carruba, G.S. Hotamisligil, Tumor necrosis factor alpha mediates apoptosis of brown adipocytes and defective brown adipocyte function in obesity, Proc. Natl. Acad. Sci. USA 97 (2000) 8033–8038.
- [33] W.P. Cawthorn, F. Heyd, K. Hegyi, J.K. Sethi, Tumour necrosis factor-alpha inhibits adipogenesis via a beta-catenin/TCF4(TCF7L2)-dependent pathway, Cell Death Differ. 14 (2007) 1361–1373.

- [34] J.B. Prins, C.U. Niesler, C.M. Winterford, N.A. Bright, K. Siddle, S. O'Rahilly, N.I. Walker, D.P. Cameron, Tumor necrosis factor-alpha induces apoptosis of human adipose cells, Diabetes 46 (1997) 1939–1944.
- [35] M. Tamai, T. Shimada, N. Hiramatsu, K. Hayakawa, M. Okamura, Y. Tagawa, S. Takahashi, S. Nakajima, J. Yao, M. Kitamura, Selective deletion of adipocytes, but not preadipocytes, by TNF-alpha through C/EBP- and PPARgamma-mediated suppression of NF-kappaB, Lab. Invest. 90 (2010) 1385–1395.
- [36] E.A. Miles, P. Zoubouli, P.C. Calder, Differential anti-inflammatory effects of phenolic compounds from extra virgin olive oil identified in human whole blood cultures, Nutrition 21 (2005) 389–394.
- [37] Z. Juranic, Z. Zizak, Biological activities of berries: from antioxidant capacity to anti-cancer effects, BioFactors 23 (2005) 207–211.
- [38] E.M. Coates, G. Popa, C.I. Gill, M.J. McCann, G.J. McDougall, D. Stewart, I. Rowland, Colon-available raspberry polyphenols exhibit anti-cancer effects on in vitro models of colon cancer, J. Carcinog. 6 (2007) 4.
- [39] A. Ejaz, D. Wu, P. Kwan, M. Meydani, Curcumin inhibits adipogenesis in 3T3-L1 adipocytes and angiogenesis and obesity in C57/BL mice, J. Nutr. 139 (2009) 919-925
- [40] H.S. Moon, H.G. Lee, Y.J. Choi, T.G. Kim, C.S. Cho, Proposed mechanisms of (-)-epigallocatechin-3-gallate for anti-obesity, Chem. Biol. Interact. 167 (2007) 85–98
- [41] S. Wolfram, Y. Wang, F. Thielecke, Anti-obesity effects of green tea: from bedside to bench, Mol. Nutr. Food Res. 50 (2006) 176–187.
- [42] Y.E. Dommels, M.M. Haring, N.G. Keestra, G.M. Alink, G.M. Alink, P.J. van Bladeren, B. van Ommen, The role of cyclooxygenase in n-6 and n-3 polyunsaturated fatty acid mediated effects on cell proliferation, PGE(2) synthesis and cytotoxicity in human colorectal carcinoma cell lines, Carcinogenesis 24 (2003) 385-392.
- [43] S. Rayalam, J.Y. Yang, S. Ambati, M.A. Della-Fera, C.A. Baile, Resveratrol induces apoptosis and inhibits adipogenesis in 3T3-L1 adipocytes, Phytother. Res. 22 (2008) 1367–1371.
- [44] K.C. Chen, L.S. Chang, Notexin upregulates Fas and FasL protein expression of human neuroblastoma SK-N-SH cells through p38 MAPK/ATF-2 and JNK/c-Jun pathways, Toxicon 55 (2010) 754–761.
- [45] H.J. Park, J.Y. Yang, S. Ambati, M.A. Della-Fera, D.B. Hausman, S. Rayalam, C.A. Baile, Combined effects of genistein, quercetin, and resveratrol in human and 3T3-L1 adipocytes, J. Med. Food 11 (2008) 773-783.
- [46] S. Rayalam, M.A. Della-Fera, J.Y. Yang, H.J. Park, S. Ambati, C.A. Baile, Resveratrol potentiates genistein's antiadipogenic and proapoptotic effects in 3T3-L1 adipocytes, J. Nutr. 137 (2007) 2668–2673.
- [47] H. Vierheilig, B. Bago, C. Albrecht, M.J. Poulin, Y. Piche, Flavonoids and arbuscular-mycorrhizal fungi, Adv. Exp. Med. Biol. 439 (1998) 9–33.
- [48] J.Y. Yang, M.A. Della-Fera, S. Rayalam, C.A. Baile, Effect of xanthohumol and isoxanthohumol on 3T3-L1 cell apoptosis and adipogenesis, Apoptosis 12 (2007) 1953–1963.
- [49] V. Mendes, R. Monteiro, D. Pestana, D. Teixeira, C. Calhau, I. Azevedo, Xanthohumol influences preadipocyte differentiation: implication of antiproliferative and apoptotic effects, J. Agric. Food Chem. 56 (2008) 11631–11637.
- [50] T.D. Gilmore, Introduction to NF-kappaB: players, pathways, perspectives, Oncogene 25 (2006) 6680–6684.
- [51] J.Y. Yang, M.A. Della-Fera, S. Rayalam, C.A. Baile, Enhanced effects of xanthohumol plus honokiol on apoptosis in 3T3-L1 adipocytes, Obesity (Silver Spring) 16 (2008) 1232–1238.
- [52] I.N. Sergeev, C.T. Ho, S. Li, J. Colby, S. Dushenkov, Apoptosis-inducing activity of hydroxylated polymethoxyflavones and polymethoxyflavones from orange peel in human breast cancer cells, Mol. Nutr. Food Res. 51 (2007) 1478–1484.

- [53] M.H. Pan, Y.S. Lai, C.S. Lai, Y.J. Wang, S. Li, C.Y. Lo, S. Dushenkov, C.T. Ho, 5-Hydroxy-3,6,7,8,3',4'-hexamethoxyflavone induces apoptosis through reactive oxygen species production, growth arrest and DNA damage-inducible gene 153 expression, and caspase activation in human leukemia cells, J. Agric. Food Chem. 55 (2007) 5081–5091.
- [54] H. Xiao, C.S. Yang, S. Li, H. Jin, C.T. Ho, T. Patel, Monodemethylated polymethoxyflavones from sweet orange (*Citrus sinensis*) peel inhibit growth of human lung cancer cells by apoptosis, Mol. Nutr. Food Res. 53 (2009) 398– 406.
- [55] I.N. Sergeev, S. Li, C.T. Ho, N.E. Rawson, S. Dushenkov, Polymethoxyflavones activate Ca²⁺-dependent apoptotic targets in adipocytes, J. Agric. Food Chem. 57 (2009) 5771–5776.
- [56] G. Swarnkar, K. Sharan, J.A. Siddiqui, B. Chakravarti, P. Rawat, M. Kumar, K.R. Arya, R. Maurya, N. Chattopadhyay, A novel flavonoid isolated from the steambark of *Ulmus wallichiana* planchon stimulates osteoblast function and inhibits osteoclast and adipocyte differentiation, Eur. J. Pharmacol. 658 (2011) 65–73.
- [57] K. Hirota, K. Morikawa, H. Hanada, M. Nonaka, Y. Nakajima, M. Kobayashi, R. Nakajima, Effect of genistein and daidzein on the proliferation and differentiation of human preadipocyte cell line, J. Agric. Food Chem. 58 (2010) 5821–5827.
- [58] S. Rayalam, M.A. Della-Fera, S. Ambati, J.Y. Yang, H.J. Park, C.A. Baile, Enhanced effects of 1,25(OH)(2)D(3) plus genistein on adipogenesis and apoptosis in 3T3-L1 adipocytes, Obesity (Silver Spring) 16 (2008) 539-546.
- [59] J.Y. Yang, M.A. Della-Fera, D.L. Hartzell, C. Nelson-Dooley, D.B. Hausman, C.A. Baile, Esculetin induces apoptosis and inhibits adipogenesis in 3T3-L1 cells, Obesity (Silver Spring) 14 (2006) 1691–1699.
- [60] J.Y. Yang, M.A. Della-Fera, C. Nelson-Dooley, C.A. Baile, Molecular mechanisms of apoptosis induced by ajoene in 3T3-L1 adipocytes, Obesity (Sliver Spring) 14 (2006) 388–397.
- [61] J.Y. Yang, M.A. Della-Fera, C.A. Baile, Guggulsterone inhibits adipocyte differentiation and induces apoptosis in 3T3-L1 cells, Obesity (Silver Spring) 16 (2008) 16–22.
- [62] H. Itokawa, Z.Z. Ibraheim, Y.F. Qiao, K. Takeya, Anthraquinones, naphthohydroquinones and naphthohydroquinone dimers from *Rubia* cordifolia and their cytotoxic activity, Chem. Pharm. Bull. (Tokyo) 41 (1993) 1869–1872.
- [63] D.Y. Jun, C.R. Han, J.Y. Lee, W. Park, M.S. Choi, M.H. Woo, Y.H. Kim, Anti-adipogenic activity of 2-carbomethoxy-2,3-epoxy-3-prenyl-1,4-naphthoquinone from *Rubia cordifolia* L., J. Med. Food 14 (2011) 454–461.
- [64] W. Guo, S. Wong, W. Xie, T. Lei, Z. Luo, Palmitate modulates intracellular signaling, induces endoplasmic reticulum stress, and causes apoptosis in mouse 3T3-L1 and rat primary preadipocytes, Am. J. Physiol. Endocrinol. Metab. 293 (2007) E576–E586.
- [65] I. Hainault, M. Carolotti, E. Hajduch, C. Guichard, M. Lavau, Fish oil in a high lard diet prevents obesity, hyperlipemia, and adipocyte insulin resistance in rats, Ann. NY Acad. Sci. 683 (1993) 98–101.
- [66] H.K. Kim, M. Della-Fera, J. Lin, C.A. Baile, Docosahexaenoic acid inhibits adipocyte differentiation and induces apoptosis in 3T3-L1 preadipocytes, J. Nutr. 136 (2006) 2965–2969.
- [67] X. Sun, M.B. Zemel, Role of uncoupling protein 2 (UCP2) expression and 1alpha, 25-dihydroxyvitamin D3 in modulating adipocyte apoptosis, FASEB J. 18 (2004) 1430–1432.
- [68] K.A. Page, D.L. Hartzell, C. Li, A.L. Westby, M.A. Della-Fera, M.J. Azain, T.D. Pringle, C.A. Baile, Beta-adrenergic receptor agonists increase apoptosis of adipose tissue in mice, Domest. Anim. Endocrinol. 26 (2004) 23–31.
- [69] H.K. Kim, M.A. Della-Fera, D.B. Hausman, C.A. Baile, Effect of clenbuterol on apoptosis, adipogenesis, and lipolysis in adipocytes, J. Physiol. Biochem. 66 (2010) 197–203.